

NOTE

GALLOCATECHIN AND TRANS SYRINGIN FROM *LIMONIASTRUM GUYONIANUM BOIS* GROWING IN TUNISIA

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ABSTRACT: Chemical investigation of butanol extract from *L. guyonianum Bois* roots led to the isolation of Gallocatechin $\underline{1}$ and Trans-Syringin $\underline{2}$. Their structures were established by means of 1 and 2D NMR spectroscopy and ES Mass spectrometry.

Key words: L. guyonianum Bois, Plumbaginaceae, roots, Gallocatechin 1, Trans Syringin 2, 1 and 2D NMR.

RESUME: des études chimiques de l'extrait butanolique des racines de l'espèce *L. guyonianum Bois*, nous ont permis d'isoler et d'identifier le Gallocatéchine <u>1</u> et le Trans-Syringine <u>2</u> signalés pour la première fois dans la plante. L'élucidation de leurs structures a été établie par le moyen de la RMN 1 et 2D confirmée par la spectrométrie de masse en mode Electro Spray.

Mots clés: L. guyonianum, Plumbaginaceae, racines, Gallocatéchine 1, Trans Syringine 2, RMN 1 et 2D.

INTRODUCTION

Limoniastrum guyonianum is a plant covered with calcareous concretions of 20 to 40 cm height, having erect branches, linear and semi-cylindrical leaves of 30 to 50mm, the sessiles are surrounding the stem [1]. The plant inflorescence is composed of Dinaric spikelet, 1-2 floras on an axis in a zig-zag. The decoction of its roots is used as depurative, galls are used for the tanning of leathers and also intervene for the tincture of hair in the Tunisian south [2], this specie is mainly distributed in the Sahara, it is known for its resistance to arid conditions. Limoniastrum guyonianum has not been reported for previous chemical investigation, therefore, bioguided fractionation of leaves extract from Limoniastrum feei endemic to Algeria led to the isolation of seven polyphenolic constituents: Gallic acid, Myriaphenone A, Myricetin-3-O- β -Galactopyranoside, Epigallocatechin gallate, Myrcetin-3-O- α -rhamnopyranoside, Quercetin and Myricetin [3]. In the course of searching new Natural products from Tunisian plants [4-8], we have examined the roots of L. guyonianum collected from Skanes, Monastir region, Tunisia and isolated for the first time from the indicated plant Gallocatechin 1 and Trans Syringin 2. Their structure identification is described in this paper.

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RESULTS AND DISCUSSION

Compound 1: The ElectroSpray Mass Spectrum of compound 1 gave pseudomolecular ions at 307 m/z [M+H] + and 329 m/z [M+Na] + in good agreement with the calculated mass for a molecular formula of $C_{15}H_{14}O_7$. ¹H NMR spectrum of compound <u>1</u> displayed a doublet appearing at δ_H 4.44 $(^{1}H, J=7.2Hz, H_{2})$ and two double doublets at δ_{H} 2.44 $(^{1}H, J_{1}=16.8 Hz, J_{2}=7.5 Hz, H_{4\alpha})$ and δ_{H} 2.65 (1 H, J₁=16.8Hz, J₂=3.0Hz, H_{4B}) which are characteristic signals of ring A from catechin nucleus (Table I). Two doublets at δ_H 5.81 and 5.85 ppm (J=2.4Hz) are assigned to H₆ and H₈ protons respectively. The observation of an additional singlet at δ_H 6.41 ppm (2H, H₂, H₆) suggested the presence of $C_{3'}$, $C_{4'}$ and $C_{5'}$ trihydroxy group substitutions in ring C. Comparison of ${}^{1}H$ and ${}^{13}C$ NMR data with those of the literature [9], suggested that compound $\underline{1}$ has a catechin skeletal pattern. Furthermore, significant long range correlations H₂-C₃; H₂-C₄; H₄-C₁₀ and H₄-C₉ confirming the presence of pyran ring having a C₃ hydroxyl group. 2D CHCorr and HMBC (300 MHz, CD₃OD) analysis showed that compound 1 was Gallocatechin. In fact, the identity of pyran ring having a C₃ hydroxyl group was confirmed by significant long range correlations H₂-C₃; H₂-C₄; H₄-C₉ and H₄-C₁₀ observed in HMBC spectrum (Figure 1). The same spectrum allowed to suggest that C aromatic ring is trisubstituted by three hydroxyl groups in positions 3', 4' and 5' on the basis of HMBC correlations H₂-C₁; H₂-C₂; H₂-C₆; H₆-C₄ and H₂-C₄. In conclusion, analysis of 1 and 2D NMR spectra compared with the literature values, allowed to propose for compound $\underline{1}$ the structure of Gallocatechin isolated for the first time from the butanol extract from L. guyonianum roots.

Table I. ¹³C NMR and ¹H NMR spectral data for compound <u>1</u>

Compound 1 (CD₃OD, 300MHz)

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Position	¹³ C	¹ H		
1		-		
2	81.5	4.44 d (J=7.2Hz)		
3	67.4	4.10 ; m		
4α	26.7	2.44; dd (J ₁ =16.8 <i>Hz</i> ; J ₂ =7.5 <i>Hz</i>)		
4β	26.7	2.65; dd (J ₁ =16.8 <i>Hz</i> ; J ₂ =3 <i>Hz</i>)		
5	155.4	-		
6	94.9	5.85; d; (J=2.4 <i>Hz</i>)		



7	156.6	-
8	98.7	5.81; d; (J= 2.4 <i>Hz</i>)
9	156.2	-
10	99.3	-
1′	132.2	-
2′	105.6	6.41; s
3′	145.3	-
4′	132.6	-
5′	145.3	-
6′	105.6	6.41; s

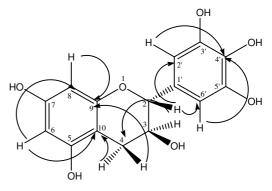


Figure 1: Heteronuclear multiple-bond correlations for compound $\underline{1}$ Arrows point from proton to carbon

Compound 2: has been isolated from *L. guyonianum* butanol extract. Its ^{1}H NMR spectrum showed a typical singlet integrating two protons appearing at δ_{H} 6.75ppm (H₅, H₉), indicating the presence of a symmetric tetra-substituted aromatic ring, another singlet at δ_{H} 3.86ppm (6H) attributable to two methoxyle groups. The observation of two characteristic doublet of triplets at δ_{H} 6.55ppm (1H, J₁=15.9Hz and J₂=2Hz, H₃) and δ_{H} 6.32ppm (1H, J₁=15.9Hz and J₂=5.4Hz, H₂) (Table II) suggested the presence of a Trans disubstituted double bond directly linked to a methylene group. The ^{13}C NMR spectrum obtained in methanol showed the famous peak of anomeric carbon C₁· at δ_{c} 105.9ppm and five peaks appearing between δ_{c} 62.9ppm and δ_{c} 78.0ppm attributable to C₂·, C₃·, C₄· and C₅· of a glycopyranose moiety and indicating that compound $\underline{2}$ is an aromatic glycoside derivative. The structure of compound $\underline{2}$ as Trans-Syringin was established on the basis of HMBC spectrum which displayed significant ^{1}H - ^{13}C long range correlations: H₅-C₆; H₅-C₇; H₅-C₃; H₉-C₃; H₂-C₁; H₉-C₈ and H₁·-C₇ (Figure 2). In conclusion, comparison of 1 and 2D NMR data of compound $\underline{2}$ with those of the literature [10-12] allowed confirming its structure of Trans-Syringin isolated for the firt time from *L. guyonianum* specie.

Table II. ¹³C NMR and ¹H NMR spectral data for compound <u>2</u> Compound 2 (CD₃OD, 300MHz)

Position	¹³ C	¹ H
1	63.9	4.22 ; dd (J ₁ =5.7 <i>Hz</i> , J ₂ =1.2 <i>Hz</i>)
2	130.5	6.32 ; dt (J ₁ =15.9 <i>Hz</i> , J ₂ =5.4 <i>Hz</i>)
3	131.7	6.55 ; d (J=15.9 <i>Hz</i>)
4	-	-
5,9	104.5	6.75; s
6,8	154.8	-



7	-	-
1'	105.8	m
2′	71.0-78.8	3.00-3.80; m
3′	71.0-78.8	3.00-3.80; m
4′	71.0-78.8	3.00-3.80; m
5′	71.0-78.8	3.00-3.80; m
6'a	62.9	3.64; dd (J ₁ =11.7 <i>Hz</i> , J ₂ =5.1 <i>Hz</i>)
6'b	62.9	3.77; dd (J ₁ =12 <i>Hz</i> , J ₂ =2.4 <i>Hz</i>)
10,11 (-OCH₃)	57.5	3.86; s

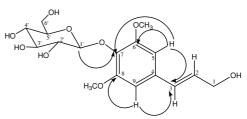


Figure 2: Heteronuclear multiple-bond correlations for compound $\underline{2}$ Arrows point from proton to carbon

EXPERIMENTAL

Plant material: *Limoniastrum guyonianum* (Plumbaginaceae) was collected in Skanes region, Monastir, Tunisia in October 2008. It was identified by Dr. F. Harzallah-Skhiri (Institut Supérieur de Biothechnologie de Monastir) and a voucher specimens were deposited in Natural Substances and organic Synthesis Laboratory, Faculty of Sciences of Monastir, Tunisia. Roots were exhaustively extracted with methanol during 5 days. Evaporation of organic solvent under reduced pressure yielded 275g of methanolic crude extract which was concentrated on a rotator evaporator under reduced pressure.

Extraction and isolation of the chemical constituents:

Compound 1: The hydro-methanolic solution has been the subject of a liquid/liquid partition using PE, EtOAc and butanol as solvents having increasing polarity. The crude butanol extract (30g) thus obtained, was chromatographed on a first silica gel column using a mixture of PE/EtOAc (4:6) gradually increased with EtOAc and Methanol as eluent to collect 247x200 mL fractions regrouped on fifteen main groups having different chemical composition (F_1 - F_{15}). The fifth one was further purified on a second silica gel column eluted with PE/EtOAc (8:2) mixture to afford 60mg of compound $\underline{1}$ as a white solid.

Compound 2: fraction F_{11} deriving from butanol extract separation has been the subject of a solid-liquid chromatographic purifications successively over two silica gel columns eluted with PE/EtOAc (7:3) mixture gradually increased with EtOAc to afford 5mg of compound **2** as a white solid.

REFERENCES

- [1] E. Floc'h, Contribution à une étude éthnobotanique de la Flore Tunisienne. Programme Flore et végétation tunisiennes. Imprimerie Officielle de la République Tunisienne, TUNISIE, **1983**.
- [2] V. Fintelmann, R. F. Weiss. Manuel pratique de Phytoterapie. Vigot, 2004, 184.
- [3] M. Chaabi, N. Beghidija, S. Benayache and A. Lobstein, Verlag der Zeitschrift für Naturforschung, Tübingen, 2008, 801.
- [4] M. A. Mahjoub, S. Ammar and Z. Mighri, Natural product Research, 2005, 19(8), 723.
- [5] H. Ben Jannet, A. Chaari, A. Bakhrouf and Z. Mighri, Natural product Research, 2006, 20(3), 299.
- [6] S. Hammami, I.Khoja, H. Ben Jannet, M. Ben Halima and Z. Mighri, J. Essent. Oil bearing plants, 2006, 9 (2), 156.
- [7] (a) H. Braham, H. Ben Jannet, Z. Mighri, *J. Soc. Chim. Tunisie*, **2007**, *9*, 109. (b) A. Bergaoui, S. Hammami, M. Ben Halima Kamel, L. Sakka Rouis, O. Boussada, D. Houas and Z. Mighri, *Journal of Enthomology*, **2008**, *5*(4), 277.
- [8] S. Hammami, N. Mighri, H. Ben Jannet, N. Boughalleb, A. Zardi-Bergaoui, A. Nefzi, P. Abreu and Z. Mighri, *Natural product Research*, **2009**, 23(16), 1466.
- [9] A. Karioti, C. Gabbiani, L. Messori, A. R. Bilia, H. Skalta, Tetrahedron Lett., 2009, 50, 1771.
- [10] E. Pretsch, P. Bühlmann, C. Affolter, Springer-Verlag Berlin Heidelberg, 2000, 97.
- [11] C. Jin, R. G. Micetich, M. Daneshtalab, Phytochemistry, 1999, 50, 677.
- [12] A. Bergaoui, S. Hammami, H. Ben Jannet, M. L. Ciavatta, G. Cimino and Z. Mighri, J. Soc. Alger. Chim., 2004, 14(2), 235.